- 1 Collective behaviour is not robust to disturbance, yet parent and offspring
- 2 colonies resemble each other in social spiders
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14 Abstract

15 Groups of animals possess phenotypes such as collective behaviour, which may determine the fitness 16 of group members. However, the stability and robustness to perturbations of collective phenotypes in 17 natural conditions is not established. Furthermore, whether group phenotypes are transmitted from parent to offspring groups is required for understanding how selection on group phenotypes 18 19 contributes to evolution, but parent-offspring resemblance at the group level is rarely estimated. We 20 evaluated robustness to perturbation and parent-offspring resemblance of collective foraging 21 aggressiveness in colonies of the social spider Anelosimus eximius. Among-colony differences in 22 foraging aggressiveness were consistent over time but changed if the colony was perturbed through 23 the removal of individuals, or via their removal and subsequent return. Offspring and parent colony 24 behaviour were correlated, but only once the offspring colony had settled after being translocated.

The parent-offspring resemblance was not driven by a shared elevation but could be due to other environmental factors. Laboratory collective behaviour was not correlated with behaviour in the field. Colony aggression seems sensitive to initial conditions and easily perturbed between behavioural states. Despite this sensitivity, offspring colonies have collective behaviour that resembles that of their parent colony, provided they are given enough time to settle into the environment.

31 Key words: Anelosimus, collective behaviour, heritability, behavioural state, perturbation

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33 Introduction

34 Many organisms form groups (Ward and Webster 2016). These aggregations can help individuals 35 avoid predation, acquire resources, find mates, and so on (Bilde et al. 2007; Frank 2007; Dobson et 36 al. 2012; Almberg et al. 2015; Groenewoud et al. 2016). For many of these purposes, groups use 37 collective behaviour, where individuals act in a co-ordinated or synchronised manner (Sumpter 38 2006). Collective behaviours cannot always be understood in terms of a simple sum of the actions of 39 individuals and so groups can possess phenotypes that simply do not exist at the individual level 40 (Parrish and Edelstein-Keshet 1999; Modlmeier et al. 2014; Farine et al. 2017). Group phenotypes 41 are therefore a tier of biological organisation that require direct study, both in terms of how they 42 relate to selection and evolution at the individual level, as well as in and of themselves (Couzin 2009). 43

Individual traits can range from being highly consistent within an individual to highly
variable (Bell et al. 2009). An individual might retain its behaviour in spite of a disturbance, or it
might find its behaviour changed as a result of a disturbance (Tuomainen and Candolin 2010; Sih et
al. 2011). The same could be true of group phenotypes; the collective behaviour of groups may resist
disturbances, or it may be altered by them (Flack et al. 2005, 2006; Smith et al. 2013; Kubitza et al.

49 2015; Formica et al. 2016). For instance, collective behaviours might be "self-organised", where 50 individuals re-create the same group behaviour after disturbances by following the same set of interaction patterns that created the initial group behaviour (Bonabeau et al. 1997; Fisher and Pruitt 51 52 2019; Fisher et al. 2019). In contrast, groups might change their behaviour following disturbances, if 53 they are shunted into different "states" following a disturbance (Flack et al. 2005, 2006; Doering et 54 al. 2018; Pruitt et al. 2018), or engage in non-linear interactions that give divergent trajectories, and 55 so different group phenotypes, from a similar set of starting conditions (May and Oster 1976; Cole 56 1994; Fisher et al. 2018; Honegger and de Bivort 2018). However, the robustness of group 57 phenotypes to disturbances is not well documented (Flack et al. 2005, 2006; Smith et al. 2013; Kubitza et al. 2015; but see: Formica et al. 2016). 58 59 If group phenotypes are resistant to disturbances and stable over time, then they can influence 60 the survival and reproductive success of individuals within those groups (Wray et al. 2011; Keiser 61 and Pruitt 2014; Pruitt and Goodnight 2014; Pruitt et al. 2017, 2019). Stability in group phenotypes 62 is important because it determines the degree to which they can be subject to natural selection (in a 63 population of groups, if all group phenotypes vary widely these phenotypes cannot be associated 64 with relative fitness). One of the most extreme forms of group disturbance is group fission, whereby 65 a subset of group members disperse or bud off to form a smaller, "daughter" group (Vollrath 1982; 66 Aviles 1986). The collective behaviour of these daughter groups can be similar to that of their parent 67 group and so exhibit a crude kind of collective or group-level heritability (Bienefeld and Pirchner 68 1990; Pruitt et al. 2017, 2019). However, unlike individual-level traits (Houle 1992), the heritability 69 of group-level traits is not widely documented. This therefore makes it hard to judge how, if at all, 70 group-level selection can contribute to evolution and adaptation (Wilson 1997b,a; Gardner and Grafen 2009; Queller and Strassmann 2009). 71

We therefore had two questions surrounding collective behaviour. First, is collective
behaviour robust to disturbance? Second, is collective behaviour transmitted from parent group to

74 offspring group in staged fission events? If both of these are true, then we might expect group 75 phenotypes such as collective behaviours to play a more important role in evolution than is currently 76 thought. We investigated these questions in a Neo-tropical social spider, Anelosimus eximius 77 (Araneae: Theridiidae). Anelosimus eximius is classified as "non-territorial permanent social" (Avilés 1997), where individuals (sometimes numbering into the 10,000s; Avilés 1997) from overlapping 78 79 generations live together in the same web structure and cooperate in web-building, prey capture, and 80 alloparental care (Vollrath 1986; Ebert 1988; Avilés and Tufiño 1998; Avilés and Harwood 2012; 81 Avilés and Guevara 2017; Pruitt and Avilés 2017). This allows them to feed on larger prey than 82 would be expected of a spider of their body size and to endure environments where related species 83 with lower levels of sociality cannot (Guevara and Avilés 2015; Avilés and Guevara 2017; 84 Fernandez-Fournier et al. 2018). Once prey make contact with the web, social spiders collectively 85 rush to immobilise it. How quickly the colony responds to a potential prey item can be an important 86 determinant of colony success and so this is the collective behaviour that we focus on here (hereafter 87 "foraging aggressiveness"; Lichtenstein et al. 2019).

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89 Methods

90 Data collection

Our study took place in June and July 2019, near Tena, Ecuador (Fig. 1), under the Ecuadorian
Ministry of the Environment permit no. 014-2019-IC-FLO-DNB/MA. We located colonies of *A*. *eximius* on roadsides, where they are relatively conspicuous on hedgerows, fences, and in trees. Their
webs are composed of a "basket" at the base, with a sheet and tangle capture web above (Yip et al.
2008). Once we found colonies, we marked their location and recorded GPS coordinates to allow us
to re-locate them. We then recorded their elevation and measured the height, width and depth of the
basket. We found 45 colonies that were suitable for our study, being within reach of an observer and

98 located within a morning's drive of our laboratory. We tested these 45 colonies' foraging 99 aggressiveness three times over six days (every other day). Our test for foraging aggressiveness was 100 the colony's speed to attack a vibrating stimulus (following: Lichtenstein et al. 2019). We stimulated 101 colonies to attack by touching a piece of wire fixed to a modified handheld vibratory device (8" 102 Vibrating Jelly Dong, Top Cat Toys, Chatsworth CA, USA) to a small piece of leaf placed in the 103 web. The leaf was always placed on the edge of the basket of the web, and we waited at least 60 104 seconds from the placement of the leaf before introducing the vibrations. The vibrations running 105 through the leaf simulate a prey item caught in the web; assays similar to this are often used to 106 estimate foraging aggressiveness in social (e.g. Laskowski and Pruitt 2014; Lichtenstein et al. 2019) 107 and solitary (Dirienzo and Montiglio 2016; Montiglio and DiRienzo 2016) spiders. We timed the 108 number of seconds from the start of the vibrations until a spider touched the leaf. If the colony did 109 not respond within 10 minutes the score was set at 600 (2.3% of all trials). This test is repeatable 110 among-colonies over four days (r = 0.26) and, at high altitudes, influences colony survival over a 11 month period (Lichtenstein et al. 2019), indicating it captures relatively stable aspects of colony 111 112 collective behaviour.

After these three baseline collective aggressiveness tests, we assigned each colony randomly 113 to one of three treatments. Fifteen colonies were "removal", 15 "procedural control" and 15 114 "control". For the removal and procedural control colonies, we returned three days after the 3rd 115 116 behavioural test and removed a subset of spiders from each colony, placed them in sealed plastic 117 boxes (190 x 190 x 90 mm) with sticks to support web building, and transported them back to our 118 laboratory. Individuals were collected either by gently shaking the web and caching spiders that 119 dropped or scooping a small bit of webbing into a large plastic box. We counted the number of 120 individuals that were large (>2mm in body length), medium sized (<2mm & >1mm in body length) 121 or small (<1mm in body length), with size being estimated by eye. We endeavoured not to destroy

any vegetation the web was built on, in order to preserve the web's structure. Control colonies were

123 left undisturbed.

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Figure 1. Maps showing the location of each of the *Anelosimus exemius* colonies in the study, with the elevation of the colony indicated by the colour (red = high elevation, blue = low elevation). In the right map the towns of Tena and Archidona are indicated with white points.

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130 Each subset of spiders that we collected was left undisturbed to acclimatise to captivity in their box 131 for two days. Boxes had four airholes to provide oxygen, and spiders were provided a moist piece of paper on the 4th day of their captivity for hydration; they were not fed. We then tested the foraging 132 133 aggressiveness of each of the 30 captive colonies three times over six days (every other day; the 1st 134 laboratory test beginning five days after the last pre-disturbance test). We modified the assay slightly 135 to account for the new setting: we reduced the power of the vibrations to avoid over-amplification in 136 the small box, and the wire was touched directly to the web rather than to a small leaf. These 137 laboratory assays were used to assess the resemblance of parent and daughter colonies in a common 138 garden environment. Although we might expect behaviour in the laboratory to differ substantially

from that in the field, due to the lack of all natural cues (but see: Boon et al. 2008; Herborn et al. 2010; Fisher et al. 2015; Yuen et al. 2016), we might still expect the ranking of colonies in terms of their foraging aggression to be similar in both the laboratory and in the field. In this case a positive correlation would be expected.

Following their 3rd test (on the same day), the spiders from procedural control colonies were 143 placed directly back into their source (parental) colony. The colonies in this treatment group 144 145 therefore lost no spiders but experienced the physical disturbance of the sampling event. Spiders 146 from the removal treatment were placed in vegetation similar to what the parent colony had built its 147 web on, but 5-10m away from the parent colony. This was designed to mimic the fission of a colony 148 and the foundation of a new colony by a subset of individuals (sociotomy), which occurs naturally in 149 A. eximius as colonies grow in size (Vollrath 1982; Venticinque et al. 1993; Avilés 1997). These 150 "bud colonies" were used to assess the heritability of colony behaviours when in the same 151 environment as their parent colony. At this point we discovered that eight of the parent colonies had 152 been destroyed by workers clearing roadsides. Two of these colonies were in the procedural control 153 group, but we could not return the previously removed spiders to a now destroyed colony, so we 154 placed these spiders into vegetation 5-10m away as bud colonies.

155 Two days after returning them to the wild, we tested the collective aggressiveness of each 156 surviving parent colony (n = 37) and each bud colony three times over six days (every other day) 157 using the same method as before. In three instances the bud colony was completely abandoned, 158 leaving 14 bud colonies (including the additional two colonies that were originally part of the 159 procedural control group) to assay for foraging aggressiveness. To evaluate the robustness of A. 160 *eximius* colonies to disturbance, we tested for a correlation between parent colonies' pre- and post-161 disturbance behaviours. We evaluated transmission of aggressiveness from parent to daughter group 162 by testing for a correlation between the pre-disturbance behaviour of parent colonies and the 163 behaviour of bud colonies in a common garden setting (the laboratory) and a natural setting (the bud

164 colony behaviours). During the three tests of the bud colony foraging aggressiveness, we observed 165 the bud colonies frequently changing position and orientation in the vegetation. We thought it was 166 likely that there was an initial "settling" phase after returning the bud colonies to the wild from captivity. Therefore, starting eight days after their 3rd test, we tested each bud colony another three 167 168 times over six days (every other day). This procedure was meant to capture bud colony behaviour 169 following a settlement period ("settled bud behaviour", the initial three tests hereafter being referred 170 to as "initial bud behaviour"). A schematic outlining the sampling regime for the study is shown in 171 Fig. 2.

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Figure 2. A schematic demonstrating our study design. In the pre-disturbance phase 45 colonies
were tested three times over six days for foraging aggressiveness. For two thirds of these colonies (in

the "removal" and "procedural control" groups) spiders were then removed to the laboratory, where they were tested three times over six days for foraging aggressiveness. Following this, spiders in the procedural control groups were returned to their original colony, while spiders in the removal groups were placed near the original colony as "bud colonies". We then tested all original colonies and all bud colonies three times over six days. Following this we tested each bud colony another three times over six days to measure "settled" behaviour.

182

183 Data analysis

184 To assess the stability of colony behaviour over time in face of the disturbance, we initially estimated 185 the phenotypic correlation (Pearson's correlations in all cases) between the log of pre-disturbance 186 foraging aggressiveness and the log of post-disturbance foraging aggressiveness, with a colony's first 187 measure pre-disturbance paired with its first measured post-disturbance, and so on. However, this 188 does not estimate the among-colony correlation between pre- and post-disturbance behaviours, 189 instead it conflates among-colony, among-date and residual variation (analogous to the "individual 190 gambit"; Brommer 2013; Dingemanse and Dochtermann 2013). To directly estimate the among-191 colony correlation between pre- and post-disturbance foraging aggressiveness, we built multivariate 192 models in MCMCglmm (Hadfield 2010) with the logs of pre-disturbance foraging aggressiveness 193 and post-disturbance foraging aggressiveness as response variables. We entered "NA" for the post-194 disturbance trials for colonies that had been destroyed. This allowed us to include their scores for the 195 pre-disturbance trials in the model, which should improve the estimate of the among-colony variance 196 in pre-disturbance foraging aggressiveness. We estimated the among-colony variances and 197 covariance between pre- and post-disturbance foraging aggressiveness, the among-date variances for 198 these traits (but no covariance as the two behaviours were never tested on the same day) and the 199 residual variances for each behaviour (but no covariance as the two behaviours were never measured

at the same time). We included the log of colony basket volume (height*depth*width), mean centred
and scaled to a variance of one, and the trial number (1-3), mean centred, as fixed effects for each
behaviour. This was done in case colony foraging aggressiveness covaried with size (Yip et al. 2008;
Pruitt et al. 2011) and in case the colonies changed their behaviour over time.

To test if the disturbed colonies changed their behaviour more than the control colonies, we estimated the raw phenotypic correlations for each of the three treatment groups. We then we fitted the multivariate model described above to each of the three treatment groups separately and compared the magnitude and distributions of the among-colony correlations. If the control group had a stronger correlation between pre- and post-disturbance foraging aggressiveness than the removal or the procedural control groups, we could conclude that the disturbance disrupted colony collective behaviour.

211 To assess the resemblance of collective behaviour between parent and offspring colonies, we 212 first estimated the phenotypic correlations between log-transformed pre-disturbance foraging 213 aggressiveness, log-transformed laboratory foraging aggressiveness, and log-transformed bud colony 214 foraging aggressiveness, associating the first pre-disturbance trial, the first laboratory trial, and the 215 first bud trial and so on. However, phenotypic correlations such as this (including those based on 216 only a single measure of parents and offspring, e.g. Pruitt et al. 2017, or those based on averages of 217 parent and offspring colony traits, e.g. Pruitt et al., 2019) conflate among- and within-colony 218 covariance, when only the former is relevant for assessing whether more aggressive parent colonies 219 have more aggressive daughter colonies (Brommer 2013; Dingemanse and Dochtermann 2013; see 220 also Niemela and Dingemanse 2018 for a discussion of the issues with using a single measure of 221 behaviour to estimate covariances). To estimate the among-colony correlation, we built multivariate 222 models in MCMCglmm, with the logs of pre-disturbance foraging aggressiveness, laboratory 223 foraging aggressiveness, and bud foraging aggressiveness as response variables. We estimated the 224 among-colony variances and covariance between these three traits. This is analogous to a parent-

offspring regression, which overestimates heritability compared to estimates from an "animal model"
(Kruuk 2004). We did not have a colony level pedigree, nor could we calculate the relatedness
among colonies by some other means. Therefore, the parent-offspring covariance we estimate here
should be taken as an upper limit for the true colony level heritability.

229 We also estimated among-date variance for each behaviour (but no covariance as the 230 behaviours were never tested on the same day) and the residual variance for each behaviour (but no 231 covariance as the behaviours were never measured at the same time). We included the log of colony 232 volume as a fixed effect for pre-disturbance behaviour, and the number of adults removed from the 233 colony and so tested in both the laboratory and as a bud colony (summing large and medium spiders, 234 so any greater than 1mm in body length) as fixed effects for laboratory and bud behaviour. This was 235 done in case colony size impacted foraging aggressiveness. These fixed effects were scaled to a 236 mean of zero and a variance of one. We also include trial number (1-3) as a fixed effect, mean 237 centred, in case the colonies changed their behaviour over time.

We estimated the raw phenotypic correlations once with the $1^{st}-3^{rd}$ tests on the bud colonies 238 (initial bud behaviour) and once with the 4th-6th tests (settled bud behaviour). We also re-fitted the 239 multivariate model using the 4th-6th tests instead of the 1st-3rd tests. If collective behaviour was 240 inherited from parent colony to offspring colony, we expected a positive among-colony correlation 241 242 between the pre-disturbance and bud behaviours. If behaviour in the laboratory reflects behaviour in 243 the field, then there would also be a positive among-colony correlation between the pre-disturbance and laboratory foraging aggressiveness. Further, if the 4th-6th tests on the bud colonies reflects settled 244 245 behaviour, we expected the among-colony correlation between pre-disturbance foraging 246 aggressiveness and the settled bud foraging aggressiveness to be stronger than the correlation 247 between pre-disturbance foraging aggressiveness and the initial bud foraging aggressiveness.

For all multivariate models we used a Gaussian error structure for each response variable, 550,000 iterations, a burn in of 50,000, and a thinning interval of 100. Priors were set to be flat and relatively uninformative, with 70% of the phenotypic variance for the logged values of each trait placed on the residual variance, 20% on the among-colony variance, and 10% on the among-date variance (following: Brommer 2017).

253

254 Results

255 Robustness to disturbance

256 Across all treatments, pre-disturbance foraging aggressiveness showed consistent differences among 257 colonies, (repeatability (r) of logged values = 0.152, credible intervals (CIs) = -0.060 to 0.348). Post-258 disturbance for aggressiveness was also consistently different among-colonies (r = 0.376, CIs = 259 0.158 to 0.555). We therefore conclude that each colony is in a relatively stable behavioural "state" 260 of a particular level of foraging aggressiveness during the six days we measured them. The 261 phenotypic correlation between pre- and post-disturbance foraging aggressiveness was significant 262 and positive (r = 0.217, t = 2.321, df = 109, p = 0.022). At the among-colony level, pre-disturbance 263 foraging aggressiveness positively covaried with post-disturbance foraging aggressiveness, although 264 the 95% CIs of the among-colony covariance overlapped zero (covariance mode = 0.167, CIs = -265 0.103 to 0.598, correlation mode = 0.547, CIs = -0.124 to 0.850). Full model results are provided in 266 the supplementary materials (Table S1). These findings suggest that colony collective behaviour is 267 stable over time.

The phenotypic correlation between pre- and post-disturbance foraging aggressiveness in the control group was quite strong and positive (Fig. 3a, r = 0.482, t = 3.483, df = 40, p = 0.001), absent in the procedural control group (Fig. 3b, r = -0.093, t = -0.567, df = 37, p = 0.574), and weakly positive but non-significant in the removal group (Fig. 3c, r = 0.151, t = 0.809, df = 28, p = 0.425).

272	At the among-colony level, for the control group, there was a positive correlation between pre- and
273	post-disturbance foraging aggressiveness (Fig. 3a, covariance mode = 0.245 , CIs = -0.240 to 1.080 ,
274	correlation mode = 0.701 , CIs = -0.177 to 0.953), no correlation at all in the procedural control
275	group (Fig. 3b, covariance mode = -0.001 , CIs = -0.621 to 0.464, correlation mode = 0.051 , CIs = -0.001 , CIs = -0.00
276	0.773 to 0.726), and a weak positive correlation in the removal group (Fig. 3c, covariance mode =
277	0.094, CIs = -0.547 to 1.086, correlation mode = 0.637, CIs = =-0.595 to 0.925). Note that the CIs of
278	all of these correlations overlap zero and hence each other. See Tables S2-4 in the supplementary
279	materials for full model results. Among-colony correlations therefore largely matched the phenotypic
280	correlations (Fig. 3a-c). These results collectively convey that perturbing colonies by removing
281	individuals disrupted colony collective behaviour, especially if the individuals were subsequently
282	returned.



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Figure 3. The relationship between the logs of pre- and post-disturbance foraging aggressiveness in the three treatment groups (a. control, b. procedural control, c. removal). Solid lines show the phenotypic correlations, while dashed lines showed the estimated among-colony correlations from the multivariate model.

289

290 Transmission of collective behaviour

291	Colonies showed consistent differences in foraging aggressiveness in the laboratory ($r = 0.282$, CIs =
292	0.080 to 0.472). Bud colonies showed a small amount consistent differences in in the initial three
293	measures of foraging aggressiveness (r = 0.082 , CIs = 0.024 to 0.332). There was no phenotypic
294	correlation between pre-disturbance foraging aggressiveness and initial bud foraging aggressiveness
295	(Fig. 4a, $r = 0.043$, $t = 0.272$, $df = 40$, $p = 0.787$) or laboratory foraging aggressiveness (Fig. 4b, $r = 0.043$, $t = 0.272$, $df = 40$, $p = 0.787$) or laboratory foraging aggressiveness (Fig. 4b, $r = 0.043$).
296	0.065, t = 0.610, df = 88, p = 0.543). Laboratory and initial bud behaviour were also not correlated
297	(Fig. 4c, $r = -0.019$, $t = -0.120$, $df = 40$, $p = 0.905$). Correlations were also absent at the among-
298	colony level (pre-disturbance & initial bud foraging aggressiveness: Fig. 4a, covariance mode =
299	0.042, CIs = -0.314 to 0.502, correlation mode = 0.143, CIs = -0.553 to 0.814; pre-disturbance &
300	laboratory foraging aggressiveness: Fig. 4b, covariance mode = 0.004 , CIs = -0.342 to 0.431 ,
301	correlation mode = 0.133 , CIs = -0.504 to 0.651 ; laboratory & initial bud foraging aggressiveness:
302	Fig. 4c, covariance mode = 0.008 , CIs = -0.462 to 0.560 , correlation mode = 0.386 , CIs = -0.631 to
303	0.779). Full model results are given in the supplementary materials (Table S5).
304	Settled bud behaviour showed consistent differences among-colonies in foraging
305	aggressiveness (r = 0.161, CIs = 0.044 to 0.464). There was a phenotypic correlation between
306	settled bud behaviour and foraging aggressiveness (Fig. 5a, $r = 0.464$, $t = 3.317$, $df = 40$, $p =$
307	0.002), but not between settled bud behaviour and laboratory foraging aggressiveness (Fig. 5b, r =
308	-0.117, t = -0.743, df = 40, p = 0.462). At the among-colony level, settled bud foraging
309	aggressiveness was positively correlated with pre-disturbance foraging aggressiveness, although
310	the CIs overlapped zero (Fig. 5a, covariance mode = 0.136 , CIs = -0.214 to 0.696, correlation
311	mode = 0.576 , CIs = -0.269 to 0.896). Laboratory foraging aggressiveness was not correlated with
312	settled bud foraging aggressiveness (Fig. 5b, covariance mode = 0.005 , CIs = -0.534 to 0.549 ,

correlation mode = 0.133, CIs = -0.675 to 0.736). Full model results are given in the

- supplementary materials (Table S6). Therefore, as for the robustness to disturbance, phenotypic
- 315 correlations matched the among-colony correlations. These results suggest that parent and
- offspring colony collective behaviours can resemble each other, but only once the offspring colony
- had settled into an environment close to that of the parental colony's.

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Figure 4. The relationship between a. pre-disturbance foraging aggressiveness and lab foraging aggressiveness, b. pre-disturbance foraging aggressiveness and initial bud foraging aggressiveness, and c. lab foraging aggressiveness and initial bud foraging aggressiveness. Point colours indicate different colonies. Solid black lines show the phenotypic correlations, while the dashed grey lines show the estimated among-colony correlations from the multivariate model.

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The volume of the colony's basket, number of adults, and trial number did not influence foraging aggressiveness in any of the models. There was some variation among days in foraging aggression, see Tables S1-6 for estimates.



Figure 5. The relationship between a. pre-disturbance foraging aggressiveness and settled bud foraging aggressiveness, and b. lab foraging aggressiveness and settled bud foraging aggressiveness. Point colours indicate different colonies. Solid black lines show the phenotypic correlations, while the dashed grey lines show the estimated among-colony correlations.

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336 Discussion

Organisms in groups can possess collective behaviours, which can be subject to selection. How 337 338 robust these collective behaviours are to disturbance, and whether they are transmitted from parent 339 groups to offspring groups, is however not well known. Here we show that the foraging 340 aggressiveness of A. eximius colonies is consistent over a period of several weeks and presumably 341 longer, given that at high elevations foraging aggressiveness can influence colony survival over 342 many months (Lichtenstein et al. 2019). Yet, colony behaviour is not robust to perturbation, 343 especially if individuals are removed from the colony and then returned. We further found that bud 344 colonies had collective behaviour that resembled that of their parent colony, but this was only

apparent once the bud colony had spent over a week settling after the translocation and was notapparent when comparing laboratory measures of the bud colony with the parent colony.

347 First, we note here that, while the all patterns we detected in the study were qualitatively 348 same at the among-colony level as at the phenotypic level, the 95% credible intervals of all among-349 colony correlations overlapped zero. From inspection of the correlation coefficients (see also Figs. 3-350 5), we can see the among-colony correlations are often stronger than the phenotypic correlations. 351 Therefore, the overlap with zero is likely due to high uncertainty, probably due to our study using 352 fewer than 50 colonies, and fewer than 20 colonies in each treatment group, rather than a small effect 353 size. We therefore take the liberty of discussing among-colony correlations that are of the same 354 strength or stronger than an equivalent and statistically significant phenotypic correlation. We do this 355 because we consider these results to represent meaningful biological trends rather than statistical 356 error.

357

358 Collective behaviour is vulnerable to disturbance

359 There were consistent differences among colonies in both pre- and post-disturbance behaviour, but 360 no covariance between pre- and post-disturbance behaviours in the procedural control and removal 361 treatment groups. This suggests that foraging aggressiveness represents a semi-stable state that a 362 colony is in, but that the colony is shifted to a different state by perturbations, as colonies did not 363 retain the same level of foraging aggressiveness when individuals were removed or when the colony 364 was disturbed by the removal and then return of individuals. Discussing populations or ecosystems 365 as "systems" that can exist in different states has a long history in ecology (May 1974; Solé and 366 Goodwin 2000). Referring to social groups in this way is less common, but interest in the utility of 367 this viewpoint is growing (Flack et al. 2005, 2006; Doering et al. 2018; Pruitt et al. 2018). Social 368 systems have previously been shown to be vulnerable to shifts from calm to antagonistic states due to

369 the removal of key individuals (Flack et al. 2005, 2006) or due to gradual heating (Doering et al. 370 2018). Here we have found that the removal of individuals combined with a physical disturbance to 371 the colony causes the colony to shift from one state of foraging aggression to another, although we 372 did not observe a general increase in aggression due to the perturbations. In fact, mean foraging 373 aggressiveness was equal in the control and removal treatment groups, and lower (longer latencies) 374 in the procedural control group. We concluded this based on comparing the intercepts for post-375 disturbance foraging aggressiveness between the models for each treatment (although note that the 376 95% credible intervals overlapped in all cases, see Tables S2-4). Instead, we have observed that a 377 colony adopts a different, yet still repeatable, behaviour to what it displayed before the disturbance. 378 As spider colonies did not return to their original foraging aggressiveness after the 379 disturbance, consistent differences in behaviour among-colonies probably do not rely on some 380 underlying stable trait of the colony (as is suggested for "pace of life syndrome" hypotheses for 381 consistent among-individual differences in behaviour; (Réale et al. 2010)). Instead, consistent 382 differences among colonies may depend on social interactions that generate positive feedback loops 383 that cause colonies to diverge in behaviour (e.g. Luttbeg and Sih 2010). Such multiplicative 384 interactions can give systems that are highly sensitive to initial conditions, and hence give variable 385 trajectories and final states (Boyce 1992; Hastings et al. 1993; Cole 1994). Therefore, following the 386 perturbation, A. eximius colonies may engage in interactions that, despite being deterministic and so 387 giving rise to consistent behaviour, nevertheless follow divergent trajectories and so do not give the 388 same behavioural trait as the colony previously possessed (Fisher et al. 2018). Interactions between 389 individual A. eximius within the colony that catalyse increased aggression could give this dynamic, 390 while interactions between the whole colony and its environment might also generate sufficient 391 feedback. Currently, our understanding of the development of A. eximius colony collective behaviour 392 is insufficient to allow us to judge the likely relative contributions of these two possibilities. 393 However, social network analysis on the distance related social spider *Stegodypus dumicola* hint that

positive feedback within colonies can cause the accentuation of individual differences within groups
(Hunt et al. 2018), raising the possibility something similar could happen in *A. eximius*.

396 Removing individuals and then adding them back to the colony (as occurred in the procedural 397 control group) completely removed any relationship between pre- and post-disturbance foraging aggressiveness. This suggests that removing individuals for a time and then returning them 398 399 destabilises collective behaviour much more than simply removing them. The returning spiders may 400 not have been recognised by their old colony-mates, and a period of antagonism may have disrupted 401 colony behaviour. Social (and subsocial) spiders are thought to discriminate between kin and non-kin 402 (Evans 1999; Bilde and Lubin 2001; Beavis et al. 2007; Schneider and Bilde 2008; Grinsted et al. 403 2011). However, A. eximius is known to accept intruders from the different colonies as well as from 404 the same colony (Pasquet et al. 1997), suggesting there would have been limited antagonism towards 405 the returning spiders. Instead, Pasquet et al. (1997) observed that the presence of an intruder 406 increases the nearest neighbour distance within a colony. This change could then influence collective 407 foraging aggressiveness. For now, we propose that the especially destabilised foraging behaviour of 408 these colonies stems from their effectively experiencing two social disturbance as opposed to just 409 one: having both lost a subset of group members and regained them, regardless of the familiarity of 410 these group members.

411

412 Parent and offspring colony collective behaviours resemble each other, but only once413 settled into the same environment

Parent colony behaviour (pre-disturbance) only covaried with bud colony behaviour once the bud colony had settled. This suggests that a group phenotype can be transmitted from parent to offspring colonies, like individual behaviours often are. However, this was only apparent over a week after the bud colony was been returned to the wild, suggesting there is an initial settling period before the bud

colony regains the collective behaviour its parent colony showed. Further, parent colony foraging
aggressiveness did not covary with laboratory foraging aggressiveness. Behaviour in the laboratory
could therefore represent a different trait to behaviour in the wild, perhaps owing to colonies'
residing in completely different environments. In short, it could be that that bud colonies were
permitted to reassume a shared environment that drives the correlation between parent colony and
bud colony (Kruuk and Hadfield 2007). If this is so, then foraging aggressiveness might itself not be
transmitted between parent and offspring groups.

425 To evaluate the possible influence of a shared environment, we need to identify an environmental variable that could drive such a parent-offspring resemblance (Kruuk and Hadfield 426 427 2007). Foraging aggressiveness in A. eximius decreases at higher elevations (Lichtenstein et al. 428 2019), and our study included colonies from 398m to 1146m above sea level (Fig.1). We tested 429 whether it was elevation that drove the parent-offspring correlation by re-fitting the model for pre-430 disturbance and settled bud foraging aggressiveness (the model also contained laboratory foraging 431 aggressiveness as a third response, but it is not important here) with the elevation of the colony 432 (mean centred and scaled to a variance of one) as a fixed effect. In this model, pre-disturbance 433 foraging aggression was lower (latencies tended to be longer) at higher elevations, although the 434 credible intervals for the effect overlapped zero (fixed effect mode = 0.392, CIs = -0.059 to 0.836), 435 but settled bud foraging aggressiveness did not change with elevation (fixed effect mode = 0.034, 436 CIs = -1.001 to 1.261). In this model the relationship between pre-disturbance and settled bud 437 foraging aggressiveness was roughly the same as in the model without elevation (covariance mode = 438 0.119, CIs = -0.212 to 0.715, correlation mode = 0.583, CIs = -0.283 to 0.892). This therefore 439 suggests that sharing the same elevation was not driving the similarity between parent and offspring 440 colonies. However, it is possible that other environmental variables are driving the resemblance.

An alternative explanation for the parent-offspring colony resemblance is that different
colonies use different but repeatable behaviour rules to assemble colony behaviour. Therefore, once

offspring colonies had settled, they were able to recreate the collective behaviour of the parent
colony. While such a dynamic suggests a colony's collective behaviour would resist a perturbation, it
may take some time for the original collective behaviour to re-establish. Notably, extra time which
was granted to the offspring colonies because of their conspicuous readjustments in the foliage, but
not to the parent colonies post-disturbance. If we had tracked the parent colonies post-disturbance for
a longer period of time, we may have seen their foraging aggressiveness return to its pre-disturbance
level.

450 The outcome of selection on collective behaviour are quite different if collective behaviour is 451 determined by an environmental variable (other than elevation) versus a directly transmitted quality 452 of the parent colony. Relatedness within A. eximius colonies is typically very high (average r = 0.92453 across four populations in Suriname, although r was estimated as 0.18 based on two nearby colonies 454 at a site in Panama; (Smith and Hagen 1996)), and so selection at the colony level could be expected 455 to give adaptation at the colony-level (Gardner and Grafen 2009; Queller and Strassmann 2009). If 456 collective behaviour is determined by the environment, then selection will most likely favour 457 colonies that best match their behaviour to the environment. In this case, changes to populations' 458 behaviour across generations is more likely to reflect changes in habitat availabilities or selection 459 acting on some aspect of colonies' habitat preferences or dispersal abilities. In contrast, if foraging 460 aggressiveness is genuinely directly passed from parent colony to offspring colony, and given at high 461 elevations we can observe selection against high foraging aggression (Lichtenstein et al. 2019), then 462 we might expect mean aggression at high elevations to decrease across generations by selection 463 acting directly on colony behaviour.

464

465 Conclusions

466 In summary, we found that the foraging aggressiveness of A. eximius colonies is relatively stable 467 over time but can be disrupted by perturbations. Returning individuals to their source colony disrupts 468 a colony's collective foraging even more than simply removing individuals from a colony. Offspring 469 colonies have collective behaviour that resembles that of their parent colony, and this does not 470 appear to be driven by a shared elevation. Instead, other forces like shared microhabitat preferences 471 or the direct transmission of colony interaction rules, genetically determined behaviours, or plastic 472 states (e.g., hunger levels, aggression levels) may drive resemblance of parent and offspring colonies. 473 Appreciating that groups possess behavioural states, and that these states may be influenced by 474 external perturbations yet still be passed from parent group to offspring group, should help us 475 understand the role of group phenotypes in ecological and evolutionary processes. 476 477 Author contributions 478 DNF, JLLL and JNP designed the study. JY acquired the permits. DNF, JLLL & RCP collected the

data. DNF analysed the data and drafted the manuscript. All authors contributed to revisions of the
manuscript and approved the final version.

481

482 Acknowledgements

- 483 We thank J. B. Barnett, H. M. Anderson and B. L. McEwen for being excellent colleagues in the
- 484 field. We also thank T. D. Swanson and the staff of the Andes and Amazon Field School at Iyarina
- for making our stay as comfortable and enjoyable as possible. Funding was provided by a Canada
- 486 150 Research Chair award to JNP. We have no conflicts of interest.

487

488 Data accessibility

All data and R code used in the analysis will be made available upon publication.

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491 References

- 492 Almberg, E. S., P. C. Cross, A. P. Dobson, D. W. Smith, M. C. Metz, D. R. Stahler, and P. J.
- 493 Hudson. 2015. Social living mitigates the costs of a chronic illness in a cooperative carnivore.
- 494 Ecol. Lett. 18:660–667.
- Aviles, L. 1986. Sex-Ratio Bias and Possible Group Selection in the Social Spider Anelosimus
 eximius. Am. Nat. 128:1–12.
- 497 Avilés, L. 1997. Causes and consequences of cooperation and permanent-sociality in spiders. Pp.
- 498 476–498 in J. C. Choe and B. J. Crespi, eds. The Evolution of Social Behavior in Insects and
- 499 Arachnids. Cambridge University Press, Cambridge.
- 500 Avilés, L., and J. Guevara. 2017. Sociality in Spiders. Pp. 188–223 in D. R. Rubenstein and P.
- 501 Abbot, eds. Comparative Social Evolution. Cambridge University Press, Cambridge.
- 502 Avilés, L., and G. Harwood. 2012. A Quantitative Index of Sociality and Its Application to Group-
- Living Spiders and Other Social Organisms. Ethology 118:1219–1229. Wiley/Blackwell
 (10.1111).
- Avilés, L., and P. Tufiño. 1998. Colony Size and Individual Fitness in the Social Spider *Anelosimus eximius*. Am. Nat. 152:403–418. The University of Chicago PressThe American Society of
- 507 Naturalists.
- 508 Beavis, A. S., D. M. Rowell, and T. Evans. 2007. Cannibalism and kin recognition in Delena
- cancerides (Araneae: Sparassidae), a social huntsman spider. J. Zool. 271:233–237. John Wiley
 & Sons, Ltd (10.1111).
- Bell, A. M., S. J. Hankison, and K. L. Laskowski. 2009. The repeatability of behaviour: a metaanalysis. Anim. Behav. 77:771–783. Elsevier Ltd.

- Bienefeld, K., and F. Pirchner. 1990. Heritabilities for several colony traits in the honeybee (Apis
 mellifera carnica). Apidologie 21:175–183. EDP Sciences.
- 515 Bilde, T., K. S. Coates, K. Birkhofer, T. Bird, A. A. Maklakov, Y. Lubin, and L. Avilés. 2007.
- 516 Survival benefits select for group living in a social spider despite reproductive costs. J. Evol.
- 517 Biol. 20:2412–2426.
- Bilde, T., and Y. Lubin. 2001. Kin recognition and cannibalism in a subsocial spider. J. Evol. Biol.
 14:959–966.
- Bonabeau, E., G. Theraulaz, J.-L. Deneubourg, S. Aron, and S. Camazine. 1997. Self-organization in
 social insects. Trends Ecol. Evol. 12:188–193. Elsevier Current Trends.
- 522 Boon, A. K., D. Réale, and S. Boutin. 2008. Personality, habitat use, and their consequences for
- survival in North American red squirrels Tamiasciurus hudsonicus. Oikos 117:1321–1328.
- 524 Boyce, M. S. 1992. Population Viability Analysis. Annu. Rev. Ecol. Syst. 23:481–506.
- 525 Brommer, J. E. 2017. Multivariate Mixed Models in R- MCMCglmm examples.
- 526 Brommer, J. E. 2013. On between-individual and residual (co)variances in the study of animal
- personality: are you willing to take the "individual gambit"? Behav. Ecol. Sociobiol. 67:1027–
 1032.
- 529 Cole, B. J. 1994. Chaos and Behaviour: The Perspective of Nonlinear Dynamics. Pp. 423–444 in L.
- A. Real, ed. Behavioral Mechanisms in Evolutionary Ecology. University of Chicago Press.
- Couzin, I. D. 2009. Collective cognition in animal groups. Trends Cogn. Sci. 13:36–43. Elsevier
 Current Trends.
- 533 Dingemanse, N. J., and N. a Dochtermann. 2013. Quantifying individual variation in behaviour:
- mixed-effect modelling approaches. J. Anim. Ecol. 82:39–54.

535	Dirienzo, N., and PO. Montiglio. 2016. Linking consistent individual differences in web str	ructure

- and behavior in black widow spiders. Behav. Ecol. 27:1424–1431. Narnia.
- 537 Dobson, F. S., V. A. Viblanc, C. M. Arnaud, and J. O. Murie. 2012. Kin selection in Columbian
- ground squirrels: Direct and indirect fitness benefits. Mol. Ecol. 21:524–531. Blackwell

539 Publishing Ltd.

- 540 Doering, G. N., I. Scharf, H. V. Moeller, and J. N. Pruitt. 2018. Social tipping points in animal
- 541 societies in response to heat stress. Nat. Ecol. Evol. 2:1298–1305. Nature Publishing Group.
- 542 Ebert, D. 1988. Behavioral asymmetry in relation to body weight and hunger in the tropical social
- 543 spider Anelosimus eximius (Araneae, Theridiidae). J. Arachnol.
- Evans, T. A. 1999. Kin recognition in a social spider. Proc. R. Soc. London. Ser. B Biol. Sci.

545 266:287–292. The Royal Society

546 Farine, D. R., A. Strandburg-Peshkin, I. D. Couzin, T. Y. Berger-Wolf, and M. C. Crofoot. 2017.

547 Individual variation in local interaction rules can explain emergent patterns of spatial

- organization in wild baboons. Proc. R. Soc. B Biol. Sci. 284.
- 549 Fernandez-Fournier, P., J. Guevara, C. Hoffman, and L. Avilés. 2018. Trait overdispersion and the
- role of sociality in the assembly of social spider communities across the Americas. Proc. Natl.
- 551 Acad. Sci. U. S. A. 115:6010–6015. National Academy of Sciences.
- Fisher, D. N., M. Brachmann, and J. B. Burant. 2018. Complex dynamics and the development of
- behavioural individuality. Anim. Behav. 138:e1–e6.
- 554 Fisher, D. N., A. James, R. Rodriguez-Munoz, and T. Tregenza. 2015. Behaviour in captivity
- 555 predicts some aspects of natural behaviour, but not others, in a wild cricket population. Proc.

556 Biol. Sci. 282:20150708.

557 Fisher, D. N., and J. N. Pruitt. 2019. Insights from the study of complex systems for the ecology and

- evolution of animal populations. Curr. Zool., doi: 10.1093/cz/zoz016.
- 559 Fisher, K., M. West, A. M. Lomeli, S. H. Woodard, and J. Purcell. 2019. Are societies resilient?
- 560 Challenges faced by social insects in a changing world. Springer International Publishing.
- 561 Flack, J. C., M. Girvan, F. B. M. de Waal, and D. C. Krakauer. 2006. Policing stabilizes construction
- of social niches in primates. Nature 439:426–9.
- 563 Flack, J. C., D. C. Krakauer, and F. B. M. de Waal. 2005. Robustness mechanisms in primate
- societies: a perturbation study. Proc. R. Soc. B Biol. Sci. 272:1091–1099.
- 565 Formica, V., C. Wood, P. Cook, and E. Brodie. 2016. Consistency of animal social networks after
- disturbance. Behav. Ecol. 28:85–93. Oxford University Press.
- 567 Frank, S. A. 2007. All of life is social. Curr. Biol. 17:R648–R650.
- Gardner, A., and A. Grafen. 2009. Capturing the superorganism: A formal theory of group
 adaptation. John Wiley & Sons, Ltd (10.1111).
- 570 Grinsted, L., T. Bilde, and P. d'Ettorre. 2011. Cuticular hydrocarbons as potential kin recognition
- 571 cues in a subsocial spider. Behav. Ecol. 22:1187–1194. Narnia.
- 572 Groenewoud, F., J. G. Frommen, D. Josi, H. Tanaka, A. Jungwirth, and M. Taborsky. 2016.
- 573 Predation risk drives social complexity in cooperative breeders. Proc. Natl. Acad. Sci. U. S. A.
- 574 113:4104–9. National Academy of Sciences.
- 575 Guevara, J., and L. Avilés. 2015. Ecological predictors of spider sociality in the Americas. Glob.
- 576 Ecol. Biogeogr. 24:1181–1191. Wiley/Blackwell (10.1111).
- Hadfield, J. D. 2010. MCMC methods for multi-response generalized linear mixed models □: The
 MCMCglmm R package. J. Stat. Softw. 33:1–22.
- 579 Hastings, A., C. L. Hom, S. Ellner, P. Turchin, and H. C. J. Godfray. 1993. Chaos in Ecology: Is

580 Mother Nature a Strange Attractor? Annu. Rev. Ecol. Syst. 24:1–33.

- 581 Herborn, K. A., R. Macleod, W. T. S. Miles, A. N. B. Schofield, L. Alexander, and K. E. Arnold.
- 582 2010. Personality in captivity reflects personality in the wild. Anim. Behav. 79:835–843.

583 Elsevier Ltd.

Honegger, K., and B. de Bivort. 2018. Stochasticity, individuality and behavior. Curr. Biol. 28:R8–
R12. Cell Press.

Houle, D. 1992. Comparing evolvability and variability of quantitative traits. Genetics 130:195–204.

587 Hunt, E. R., B. Mi, C. Fernandez, B. M. Wong, J. N. Pruitt, and N. Pinter-Wollman. 2018. Social

interactions shape individual and collective personality in social spiders. Proc. R. Soc. B Biol.
Sci. 285.

590 Keiser, C. N., and J. N. Pruitt. 2014. Personality composition is more important than group size in

determining collective foraging behaviour in the wild. Proc. Biol. Sci. 281:20141424.

592 Kruuk, L. E. B. 2004. Estimating genetic parameters in natural populations using the "animal

593 model." Philos. Trans. R. Soc. Lond. B. Biol. Sci. 359:873–90. The Royal Society.

Kruuk, L. E. B., and J. D. Hadfield. 2007. How to separate genetic and environmental causes of
similarity between relatives. J. Evol. Biol. 20:1890–1903. Blackwell Publishing Ltd.

596 Kubitza, R. J., J. Suhonen, and T. Vuorisalo. 2015. Effects of experimental perturbation of group

structure on hierarchy formation and behaviour in House Sparrows. Ornis Fenn. 92:157–171.

- 598 Laskowski, K. L., and J. N. Pruitt. 2014. Evidence of social niche construction: persistent and
- repeated social interactions generate stronger personalities in a social spider. Proc. Biol. Sci.
 281:20133166.
- Lichtenstein, J. L. L., D. N. Fisher, B. L. McEwen, D. T. Nondorf, E. Calvache, C. Schmitz, J.

- 602 Elässer, and J. N. Pruitt. 2019. Collective aggressiveness limits colony persistence in high but
- not low elevation sites in Amazonian social spiders. bioRxiv, doi: 10.1101/610436.
- Luttbeg, B., and A. Sih. 2010. Risk, resources and state-dependent adaptive behavioural syndromes.
- 605 Philos. Trans. R. Soc. Lond. B. Biol. Sci. 365:3977–3990.
- 606 May, R. M. 1974. Biological Populations With Nonoverlapping Generations Stable Points, Stable
- 607 Cycles, And Chaos. Science (80-.). 186:645–647.
- May, R. M., and G. F. Oster. 1976. Bifurcations and Dynamic Complexity in Simple Ecological
- 609 Models. Am. Nat. 110:573–599.
- 610 Modlmeier, A. P., C. N. Keiser, J. V. Watters, A. Sih, and J. N. Pruitt. 2014. The keystone individual
- 611 concept: an ecological and evolutionary overview. Anim. Behav. 89:53–62.
- 612 Montiglio, P.-O., and N. DiRienzo. 2016. There's no place like home: the contribution of direct and
- extended phenotypes on the expression of spider aggressiveness. Behav. Ecol. 27:arw094.
- 614 Niemela, P. T., and N. J. Dingemanse. 2018. On the usage of single measurements in behavioural

ecology research on individual differences. Anim. Behav. 145:99–105.

- 616 Parrish, J. K., and L. Edelstein-Keshet. 1999. Complexity, pattern, and evolutionary trade-offs in
- animal aggregation. Science 284:99–101. American Association for the Advancement ofScience.
- Pasquet, A., M. Trabalon, A. G. Bagnères, and R. Leborgne. 1997. Does group closure exist in the
 social spider Anelosimus eximius 2? Behavioural and chemical approach. Insectes Soc. 44:159–
 169. Birkhäuser Verlag.
- Pruitt, J. N., and L. Avilés. 2017. Social spiders: mildly successful social animals with much
 untapped research potential. Anim. Behav. Academic Press.

- 624 Pruitt, J. N., A. Berdahl, C. Riehl, N. Pinter-Wollman, H. V Moeller, E. G. Pringle, L. M. Aplin, E. J.
- 625 H. Robinson, J. Grilli, P. Yeh, V. M. Savage, M. H. Price, J. Garland, I. C. Gilby, M. C.
- 626 Crofoot, G. N. Doering, and E. A. Hobson. 2018. Social tipping points in animal societies.
- 627 Proceedings. Biol. Sci. 285:20181282. The Royal Society.
- 628 Pruitt, J. N., and C. J. Goodnight. 2014. Site-specific group selection drives locally adapted group
- 629 compositions. Nature 514:359–362. Nature Research.
- 630 Pruitt, J. N., C. J. Goodnight, and S. E. Riechert. 2017. Intense group selection selects for ideal group
- compositions, but selection within groups maintains them. Anim. Behav. 124:15–24. Academic
 Press.
- Pruitt, J. N., G. Iturralde, L. Avilés, and S. E. Riechert. 2011. Amazonian social spiders share similar
 within-colony behavioural variation and behavioural syndromes. Anim. Behav. 82:1449–1455.
- 635 Academic Press.
- 636 Pruitt, J. N., B. L. McEwen, S. T. Cassidy, G. M. Najm, and N. Pinter-Wollman. 2019. Experimental
- evidence of frequency-dependent selection on group behaviour. Nat. Ecol. Evol. 3:702–707.
- 638 Nature Publishing Group.
- Queller, D. C., and J. E. Strassmann. 2009. Beyond society: The evolution of organismality. Philos.
 Trans. R. Soc. B Biol. Sci. 364:3143–3155. The Royal Society.
- 641 Réale, D., D. Garant, M. M. Humphries, P. Bergeron, V. Careau, and P.-O. Montiglio. 2010.
- 642 Personality and the emergence of the pace-of-life syndrome concept at the population level.
- 643 Philos. Trans. R. Soc. Lond. B. Biol. Sci. 365:4051–4063.
- 644 Schneider, J. M., and T. Bilde. 2008. Benefits of cooperation with genetic kin in a subsocial spider.
- 645 Proc. Natl. Acad. Sci. U. S. A. 105:10843–6. National Academy of Sciences.
- 646 Sih, A., M. C. O. Ferrari, and D. J. Harris. 2011. Evolution and behavioural responses to human-

- 647 induced rapid environmental change. Evol. Appl. 4:367–387. Blackwell Publishing Ltd.
- 548 Smith, C. E., B. J. Hurley, C. N. Toms, A. D. Mackey, M. Solangi, and S. A. Kuczaj. 2013.
- 649 Hurricane impacts on the foraging patterns of bottlenose dolphins Tursiops truncatus in

650 Mississippi sound. Mar. Ecol. Prog. Ser. 487:231–244.

- 651 Smith, D. R., and R. H. Hagen. 1996. Population structure and interdemic selection in the
- cooperative spider Anelosimus eximius. J. Evol. Biol. 9:589–608. John Wiley & Sons, Ltd
 (10.1111).
- Solé, R. V., and B. Goodwin. 2000. Signs of Life: How Complexity Pervades Biology. 1st ed. Basic
 Books, New York.
- Sumpter, D. J. T. 2006. The principles of collective animal behaviour. Philos. Trans. R. Soc. Lond.
 B. Biol. Sci. 361:5–22.
- Tuomainen, U., and U. Candolin. 2010. Behavioural responses to human-induced environmental
 change. Biol. Rev. 86:640–657. Wiley/Blackwell (10.1111).
- 660 Venticinque, E. M., H. G. Fowler, and C. A. Silva. 1993. Modes and Frequencies of Colonization
- and its Relation to Extinctions, Habitat and Seasonality in the Social Spider Anelosimus

Eximius in the Amazon (Araneidae: Theridiidae). Psyche (New York) 100:35–41.

- Vollrath, F. 1982. Colony Foundation in a Social Spider. Z. Tierpsychol. 60:313–324. John Wiley &
 Sons, Ltd (10.1111).
- Vollrath, F. 1986. Eusociality and extraordinary sex ratios in the spider Anelosimus eximius
- 666 (Araneae: Theridiidae). Behav. Ecol. Sociobiol. 18:283–287. Springer-Verlag.
- 667 Ward, A., and M. Webster. 2016. Sociality: The Behaviour of Group-Living Animals. Springer.
- 668 Wilson, D. S. 1997a. Altruism and organism: Disentangling the themes of multilevel selection

- 669 theory. P. *in* American Naturalist.
- 670 Wilson, D. S. 1997b. Introduction: multilevel selection theory comes of age. Am. Nat. 150
- 671 Suppl:S1–S21. The University of Chicago Press.
- 672 Wray, M. K., H. R. Mattila, and T. D. Seeley. 2011. Collective personalities in honeybee colonies
- are linked to colony fitness. Anim. Behav. 81:559–568.
- 674 Yip, E. C., K. S. Powers, and L. Avilés. 2008. Cooperative capture of large prey solves scaling
- 675 challenge faced by spider societies. Proc. Natl. Acad. Sci.
- 676 Yuen, C. H., N. Pillay, M. Heinrichs, I. Schoepf, and C. Schradin. 2016. Personality traits are
- 677 consistent when measured in the field and in the laboratory in African striped mice (Rhabdomys
- pumilio). Behav. Ecol. Sociobiol. 70:1235–1246. Springer Berlin Heidelberg.